



A REVIEW ON MUTATION BREEDING FOR THE IMPROVEMENT OF CROP PLANTS:
PRESENT AND FUTURE.

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ABSTRACT

Mutation, nowadays, has become an established tool in plant breeding and is generally used for developing crop cultivar/s or improving certain specific traits. An extensive work has been done for improving crop species by inducing mutations signifying their economic values. In the present work, two forms of mutation breeding i.e. natural mutations and induced mutations have been discussed. The natural mutations occur spontaneously and known to alter the structure of chromosomes in all plants, hence are harmful. While, induced mutations are generally grouped into two broad categories: chemical mutagens and physical mutagens. The physical mutagenesis is caused by UV rays, X-rays, cosmic rays, gamma rays, etc. and chemical mutagenesis is caused by EMS, colchicine, sodium azide, MNU, hydroxylamine hydrochloride, etc. Such genetic transformation by mutation breeding also creates morphological variants that act as a tool for understanding the physiology of the plants and optimistically, lead to their domestication. Presently, a number of mutants (chemicals and radiation) are being used to explore genomics, produce saturated genetic maps, for gene expression and regulation. Thus, mutation based breeding has been to renovate the well adapted plant varieties by attending one or two major trait/s even in *in vitro* culture, which limit their productivity or enhance their quality value.

Table 1: The list of mutant crops produced by X ray treatment.

Name of the crop	Traits improved	Authors
<i>Glycine max</i>	Increased oleic acid content	Rahman <i>et al.</i> 1994
<i>Vigna unguiculata</i>	Significant enhancement in length and fresh weight of shoot and leaf area	Ikram <i>et al.</i> 2015
<i>Vigna radiata</i>	<i>Fusarium</i> spp., <i>Rhizoctonia solani</i> and <i>Macrophomina phaseolina</i> were completely suppressed	Ikram <i>et al.</i> 2015
<i>Triticum</i>	Higher tolerance to adverse effects of saline condition	Asghari <i>et al.</i> 2013
<i>Corchorus capsularis</i>	Increase plant height	Sinhamahapatra and Rakshit 1990

Introduction:

Mutations play an important role in the evolution and considered as the chief source of origin of a new species or the creation of new genetic variation that the used in breeding programs. During the past few decades more than 2500 varieties have been released worldwide, which are derived either as direct mutants or from their progenies (Kharkwal *et al.* 2008). The prime strategy in mutation based breeding is to upgrade the well-adapted plant varieties by altering one or two major traits, which limit their productivity or increase their quality value. Further, the developed mutants can

be used for morphological evaluation, molecular analysis, and even in studying phylogeny.

Presently, a number of mutants induced by chemical/s and radiation are being used to produce saturated genetic maps and explore genomics, gene expression and gene regulation (Ahloowalia and Maluszynski 2001). Several morphological and other mutant/s of the economic importance and academic interests have been isolated in different crops by various researchers and the work on mutation breeding has been reviewed (Kharkwal 1983; Khan and Farook 1998; Dixit *et al.* 2000). Thus, keeping in mind the successive use of mutational breeding,

the present work has been compiled to understand the past and future of mutation breeding in crop plants.

Table 2: The list of various mutant crops produced by Gamma rays.

Name of crop plant	Traits	Authors
Legumes	Salt tolerant	Kumar <i>et al.</i> 2017
<i>Carthamus tinctorius</i>	Increase genetic variability	Kumar and Srivastva 2010
<i>Abelmoschus moschatus</i>	High yield	Warghat <i>et al.</i> 2011
<i>Polianthes tuberosa</i>	Higher genetic variability	Navabi <i>et al.</i> 2016
<i>Arachis hypogaea</i>	Increase grain yield and other morpho-agronomic parameters	Lukanda <i>et al.</i> 2013
<i>Glycine max</i>	Increase the genetic diversity and tolerance to drought	Aminah <i>et al.</i> 2015

Table 3: The list of various mutant crops produced by Colchicine treatment:

Name of plant species	Traits	Authors
Banana	Resistance to disease	Hamill <i>et al.</i> 1992
Pineapple	Resistance to disease, resistance to herbicides and antibiotics	Brar and Jain 1998
Kiwifruit	Increase fruit size	Wu <i>et al.</i> 2011
<i>Brassica napus</i>	Homozygous double haploid	Weber <i>et al.</i> 2004
<i>Helianthus annuus</i>	Rust resistance, height and maturity	Downes and Marshall 1983
Azuki bean	Increase seedling rate and plant survival rate	Pu <i>et al.</i> 2005
<i>Sorghum bicolor BT×623</i>	Longer leaf length and stronger root system	Murali <i>et al.</i> 2013
<i>Raphanus sativus</i>	Higher yield and wider adaptability	Manawadu <i>et al.</i> 2016
<i>Prunella vulgaris</i>	Superior tetraploid	Kwon <i>et al.</i> 2014

Types of Mutation Breeding:

The process mutagenesis involves sudden heritable change/s in the genetic information of an organism, not caused by genetic segregation or genetic recombination, but induced by chemical, physical or biological agents. Further mutagenesis may be categorized as: Natural and Induced mutations

1. **Natural mutations:** Such mutations occur spontaneously in all plants, by changing the chemistry of the plant genetic material or altering the structure of a chromosome. Most often the mutation is harmful and the change is erased by natural selection. Normally, they are harmful or developing new crop cultivar/s and for changing the plan of traits caused by cosmic rays, which bombard the earth constantly, can penetrate matter readily. If one strikes a chromosome it can produce a mutational change/s. Normal physical

phenomena, such as heat can cause mutations; so can pure oxygen under pressure. It has been shown that genetic material can mutate simply through the process aging. Natural evolution is based on three main factors: spontaneous mutations; hybridization, which reshuffles mutations into almost unlimited genetic patterns; and natural selection, which favours the perpetuation of individuals with genetic patterns which make them better able to survive and to reproduce.

Natural mutation happens randomly and therefore not easily traceable. There are various factors that cause these kind of mutations or increase their frequency. This includes the activity of mutagens found in nature, such as solar radiation or reactive chemicals such as depurinators or free radicals. Alterations during the replication of DNA during mitosis or unequal crossing-over events during meiosis can go un-

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repaired in a cell, leading to mutant progeny cells. It is difficult to find out that at what rate these mutations can occur, because the cellular machinery typically catches these mistakes. Occasionally, however, some alterations make it permanently into the organism's genome.

2. Induced mutations: On the basis of type of mutagen used, induced mutations are of two types: physical mutagens and chemical mutagens.

a. Physical mutagens: In the past 80 years, physical mutagens, mostly ionizing radiations, have been used widely for inducing hereditary aberrations and more than 70% of mutant varieties were developed using physical mutagenesis (Mba *et al.* 2013). Mutagenic radiations includes X-rays, ionizing particles and ultraviolet rays. The ionizing components of the Electromagnetic spectrum, include cosmic, gamma (γ) and X-rays (Mba *et al.* 2013). The most commonly used physical mutagens are X-rays. Since then, the various subatomic particles (neutrons, protons, beta particles and alpha particles) have been generated using nuclear reactors. Gamma radiations are mostly emitted by radioactive cobalt (^{60}Co) is mostly used (Acquaah 2006). However, it can be used for irradiating the whole plant and delicate materials, like pollen grains. Several mutant varieties have been developed through gamma radiation (Mba *et al.* 2010).

Ionizing emission goes deeper into the tissue and cause a great number of variations in the chemical composition. The major advantage of using physical mutagenesis compared to chemical mutagenesis is the degree of accuracy and sufficient reproducibility. During the past two decades, ion beams either through implantation or irradiation have become a new type of physical mutagen instead of the widely used gamma rays, X-rays and neutrons. The positively charged ions are accelerated at a high speed (about 20%–80% of the speed of light) and form high linear energy transfer (LET) radiation. LET radiation causes significant biological effects, such as chromosomal aberration,

lethality, etc. as compared to other types of radiation used in physical mutagenesis. The damage caused by ion beams to DNA double strands is less repairable as compared to that induced by gamma rays due to deletion of DNA fragments of various sizes. More recently, to study the intricacies of mutation induction in space, plant materials have been sent out into aerospace. It has been speculated that the special environment of space flight, such as cosmic radiation, microgravity, weak geomagnetic field, etc. contains the potential agents of mutation induction. However, knowledge of the underlying genetics of aerospace mutagenesis is so far scarce (Mba *et al.* 2012).

Ionizing radiations cause mutations by breaking chemical bonds in the DNA molecule, deleting a nucleotide, or substituting it with a new one also pointed out the importance of radiation being applied at the proper dose, a factor that depends on radiation intensity and duration of exposure. The exposure may be chronic (continuous low dose administered for a long period) or acute (high dose over a short period). The quality of mutation is not necessarily positively correlated with the dose rate. It is common knowledge that a high dose does not necessarily yield the best results. The mutagen dose used should be a compromise between a mutation load and the chance to find desirable mutations, and this greatly depends on the cost effectiveness of selection. Screening of larger mutant populations that originate from a lower mutagen dose may be feasible for traits with simple phenotypic selection criteria, such as early maturity. On the other hand, screening the same population for complex phenotypic traits, such as seed protein quality would not be feasible.

i. X rays: The first crop species to be mutagenized was barley by Stadler. In case of mutagenic barley, the resulting plants were white, yellow, pale yellow and some had white stripes. There are two novel approaches to barley mutagenesis. The most powerful approach is the use of neutron radiation for the production of deletion libraries. With this method, deletion in

the range of 100-10,000bp can be generated. The second approach deals with transposons mutagenesis (Stadler *et al.* 1930).

ii. UV rays: The mutagenic effect of ultraviolet light was discovered by Altenbung (1934) through irradiation of the polar cap cells of fruit fly eggs. In those organisms, germ tissue could be easily exposed to the low-penetrating ultraviolet light which resulted in covalent dimerization of adjacent pyrimidine. Emission of UV light (250–290 nm) has a modest capacity to infiltrate tissues as compared with ionizing radiation. The UV rays mutants were successfully produced in few crops (Table 1).

iii. Gamma rays: Gamma rays have a shorter wavelength and therefore, possess more energy

than protons. Few important improved varieties using gamma radiations are listed in the table 2.

iv. Neutrons: Neutrons are hazardous and hence have less penetrating abilities, but they are known to cause serious damage to the chromosomes. They are best used for materials, such as dry seeds. Various forms of neutrons were also studied extensively for their use in mutagenesis in the 1960s and 1970s. Though it has been proved to be an effective mutagen, particularly for producing large DNA fragment deletions, the application of neutrons in induced mutagenesis is limited. In *Capsicum annuum* higher yield is obtained by treating with neutrons (Falusi *et al.* 2012)

Table 4: The list of various mutant crops produced by sodium azide (NaN₃) treatment:

Name of plant species	Traits	Authors
<i>Hordeum vulgare</i>	Chlorophyll mutant	Prina and Fevret 1983
<i>Pisum sativum</i>	Pyridoxin deficient	Kumar 1988
<i>Musa</i> spp.	Resistant against <i>Fusarium oxysporum</i> f. sp. Cubense	Bhagwat and Duncane 1988
<i>Hordeum vulgare</i>	Anthocyanins and proanthocynidins deficient	Olsen <i>et al.</i> 1993
<i>Lactuca sativa</i>	Down mildew resistant	Okubara <i>et al.</i> 1994
<i>Avena strigosa</i>	Disease resistant	Papadopoulou <i>et al.</i> 1999
<i>Triticum aestivum</i>	Salt tolerance	Agata <i>et al.</i> 2001
<i>Oryza sativa</i>	Reduced amylase content	Jeng <i>et al.</i> 2003
<i>Oryza sativa</i>	Auxin resistant mutant	Chhun <i>et al.</i> 2003
<i>Hordeum vulgare</i> (barley)	Mildew resistant	Molina-Cano <i>et al.</i> 2003
<i>Vigna radiata</i> L.	Quantitative traits	Samiullah <i>et al.</i> 2004
<i>Lagerstroemia indica</i>	Resistant to powdery mildew and leathery foliage	White and Carl 2004
<i>Vigna radiata</i> L.	Quantitative traits	Samiullah <i>et al.</i> 2004
<i>Spathoglottis plicata</i>	Improved floricultural significance	Roy and Biswas 2005
<i>Oryza sativa</i>	Enhanced yield	Jeng <i>et al.</i> 2006
<i>Zea mays</i>	Resistant against pathogen <i>Striga</i>	Kiruki <i>et al.</i> 2006
<i>Arachis hypogaea</i>	Yield traits	Mensah and Obadoni 2007
<i>Saccharum officinarum</i>	Red rot (<i>Colletotricum falcatum</i>) resistant	Ali <i>et al.</i> 2007
<i>Arachis hypogea</i>	Disease resistant	Mondal <i>et al.</i> 2007
<i>Oryza sativa</i> (rice)	Silicon deficient mutant	Nakata <i>et al.</i> 2008
<i>Halianthus annuus</i>	Reduced triacylglycerol	Venegas-Caleron <i>et al.</i> 2008
<i>Halianthus annuus</i>	Enhanced stearic acid content	Skoric <i>et al.</i> 2008
<i>Oryza sativa</i> L.	Enhanced amylase content	Suzuki <i>et al.</i> 2008
<i>Zea mays</i>	Drought tolerant mutant	He <i>et al.</i> 2009
<i>Phaseolus vulgaris</i>	Higher antioxidant activity	Jeng <i>et al.</i> 2010

b. Chemical mutagenesis: The chemical mutagenic agents have the main advantage that they can be applied without complicated equipment or facilities. For chemical mutagens the ratio of mutational to undesirable

modifications is generally higher than for physical mutagens. For inducing mutations the material is soaked in a solution of the mutagen. During the application of chemical mutagens, the care should be taken as it is carcinogenic. Material and safety data sheets for the specific

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chemical mutagen should be chosen carefully and after the use of chemical agent it should be appropriately inactivated before disposal. From the large number of mutagenic compounds, only a few numbers have been tested in plants. Among them, only a few group of alkylating agents has shown large application in plant experimental mutagenesis and plant mutation breeding. About 80% of the registered new mutant plant varieties reported in the International Atomic Energy Association (IAEA) *database* achieved via chemical mutagenesis were induced by alkylating agents. Of these, there are compounds namely ethyl methane sulphonate (EMS), 1-methyl-1-nitrosourea which account for about 64% of these varieties are (Leitao *et al.* 2012).

Alkylating agents are common among a large group of classes of compounds, including sulphur mustards, nitrogen mustards, epoxides, ethyleneimines, ethyleneimides, alkyl methanesulphonates, alkylnitrosoureas, alkylnitrosoamines, alkylnitrosoamides, alkyl halides, alkyl sulphates, alkyl phosphates, chloroethylsulphides, chloroethylamines, diazoalkanes, etc. Alkylating agent is one of the most effective chemical mutagenic group (these agent react with the DNA by alkylating the phosphate groups as well as the purines and pyrimidines). Second group is that of the base analogues (they are closely related to the DNA bases and can be wrongly incorporated during replication) e.g. 5-bromouracil and maleic hydrazide. The point mutations created by chemical mutagens have great advantage to generate not only loss-of-function but also gain-of-function phenotypes, if the mutation leads to a modified protein activity or affinity, like tolerance to the herbicide glyphosate or sulphonylurea shown in *Medicago truncatula*. The effect of efficiency of mutagen is effected by the concentration of mutagen, the length of treatment and the temperature at which the experiment is carried out.

i. Colchicine: Colchicine is an alkaloid from *Colchicum autumnale*, which is widely used for the induction of polyploidy plants. It is the most effective and the most widely used

treatment for chromosome doubling. It has been used with great success in a large number of crop species belonging to both dicot and monocot groups (Table 3).

ii. Sodium azide: It is well known that sodium azide induces chromosomal aberrations only at a very low rate compared to other mutagenic treatments. Sodium azide was induced for the first time in Barley (Gruszka *et al.* 2012) for creating mutants, and after that, a number of crops have been produced by sodium azide induced mutation (Table 4).

iii. EMS: Ethyl methanesulphonate (EMS) is a stable and effective chemical mutagen (Table 5).

iv. MNU: A chemical, 1-methyl-3-1-nitrosoguanidinenitro, is carcinogenic, but also used as a mutagen in breeding programmes (Table 6).

v. Hydroxylamine hydrochloride: Hydroxylamine can be used to mutagenize purified DNA. It results in C to T and G to A transition mutations when double stranded DNA is mutagenized. Hydroxylamine decomposes in hot water and can be neutralized in this way. It also decomposes in atmospheric moisture and carbon dioxide and should be stored desiccated at room temperature (Table 7).

A compilation of all the information gathered from different research articles published revealed that about a large number of crops varieties, like cereals (350 varieties), Legumes (62 varieties), Fruits (40 varieties), and about 462 varieties of ornamentals plants have been produced globally. Similarly, in India, 24 varieties of Rice, 12 of Barley, and 8 of cotton and groundnut have been produced as a result of mutational breedings.

In spite of successful implementation of induced mutations in 20th century, it was removed from the main stream of crop improvement programme. A number of breeding programme have been declared closed in the last two or three decades, which affects the general crop improvement programme. However, it has

become a mainstream crop improvement method once again and significant efforts have been made to invest in few interventions. These efforts include: to promote new plant breeders, allotment of new research projects, and considerable investment in training and infrastructure. Besides, induced mutation, there

is also a need for the use of cell and molecular biology techniques to enhance/ or better understanding the mutational events. Further, in the ongoing induced mutation, the use of pre-breeding and integration of phenomics, are also suggested for the improvement of crops.

Table 5: Various mutant crops produced by EMS treatment.

Name of plant species	Trait	Authors
<i>Vigna radiata</i>	Highest callus proliferation.	Rafiq <i>et al.</i> 2012
<i>Gossypium hirsutum</i>	Drought tolerance	Witt <i>et al.</i> 2016
<i>Cucumis sativus</i>	Homozygous mutant lines	Wang <i>et al.</i> 2014
<i>Oryza sativa</i>	Lethal dose determination	Talebi <i>et al.</i> 2012
<i>Linum usitatissimum</i>	High yielding variety	Rafiq <i>et al.</i> 2017
<i>Ipomoea batatas</i>	Salt tolerant	Luan <i>et al.</i> 2007
<i>Trigonella foenum-graecum</i>	Higher seed yield	Basu <i>et al.</i> 2008

Table 6: Various mutant crops produced by MNU treatment

Name of the crop plant	Traits	Author
Rice	Leaf emergence	Suzuki <i>et al.</i> 2008
Rice	High Glutelin content	Satoh <i>et al.</i> 1979
Barley	High chlorophyll content	Kurowska <i>et al.</i> 2012

Table 7: Various mutant crops produced by hydroxylamine hydrochloride treatment

Name of the crop plant	Traits	Author
<i>Phaseolus vulgaris</i>	Change in number, aerial biomass and pods per plant.	Mosisa <i>et al.</i> 2014
<i>Sesamum indicum</i>	Plant height and internode length increased	Birara <i>et al.</i> 2014

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References:

Acquaah G (2006). Principles of plant genetics and breeding. Chichester: Wiley-Blackwell. pp: 740.

Agata R, Mario R, Linda M, Cristiano P, Giuseppe N and Natale DF (2001). Enhanced osmo-tolerance of a wheat mutant selected for potassium accumulation. *Plant science* 160: 441-448.

Ahloowalia BS and Maluszynski M (2001). Induced mutations-a new paradigm in plant breeding. *Euphytica* 118: 167-173.

Ali A, Naz S, Alam S and Iqwal J (2007). In vitro induced mutation for screening of red rot

(*Colletotrichum falcatum*) resistance in sugarcane (*Saccharum officinarum*). *Pakistan Journal of Botany* 39 (6): 1979-1994.

Altenbung E (1934). The artificial production of mutations by ultraviolet light. *American Naturalist* 68: 491-501.

Aminah, Nur A, Abdullah, Tahir N, Edy and Nuraeni (2015). Improving the genetic diversity of soybean seeds and tolerance to drought irradiated with gamma rays. *International Journal of Current Research and Academic Review* 3(6): 105-113.

Asghari R, Razavi A, Bakhtiari S and Soleymanifard S (2013). Germination of X-ray treated wheat seeds in saline conditions. *International Journal of Agriculture and Crop Sciences* 6(16): 1153-1163.

Basu SK, Acharya SN and Thomas JE (2008). Genetic improvement of fenugreek (*Trigonella foenum-graecum* L.) through EMS induced

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- mutation breeding for higher seed yield under western Canada prairie conditions. *Euphytica* 160: 249–258.
- Bhagwat B and Duncan EJ (1988). Mutation breeding of banana cv. Highgate (*Musa* spp., AAA Group) for tolerance to *Fusarium oxysporum* f. sp. *cubense* using chemical mutagens. *Scientia Horticulturae* 73: 11-22.
- Birara A, Muthuswami M and Andargie M (2014). Effect of Chemical Mutation by Sodium Azide on Quantitative Traits Variations in *Sesamum indicum* L. *Plant Sci. Today*.1(1): 33-38.
- Brar DS and Jain SM (1998). Somaclonal variation: mechanism and applications in crop improvement. In: S. M. Jain et al. (Eds). Somaclonal variation and induced mutations in crop improvement. pp. 15-37. Kluwer Academic Publishers, Dordrecht.
- Chhun T, Taketa S, Tsurumi S and Ichii M (2003). Interaction between two auxin resistant mutants and their effects on lateral root formation in rice (*Oryza sativa* L.). *Journal of Experimental Botany* 393: 2701-2708.
- Dixit GP, Tripathi DP, Chandra S, Tewari TN and Tickoo JL (2000). MULLaRP crops: varieties developed during the last fifty years. All India Coordinated Research Project on MULLaRP (ICAR), Indian Institute of Pulses Research, Kanpur, India.
- Downes RW and Marshall DR (1983). Colchicine variants in sunflower. *Euphytica*. p. 757-766.
- Falusi OA, Daudu OAY, Jaime A and Silva TD (2012). Effects of Fast Neutron Irradiation on Agronomic Traits of *Capsicum annum* (Nigerian pepper). *European Journal of Horticultural Science* 77: 41–45.
- Gruszka D, Szarejko I and Maluszynski M (2012). Sodium Azide as a mutagen. In: Shu QY, Forster BP, Nakagawa H, (eds), *Plant Mutation Breeding and Biotechnology*, pp 159-166.
- Hamill SD, Smith MK and Dodd WA (1992). *In vitro* Induction of Banana Auto-tetraploids by Colchicine Treatment of Micro propagated Diploids, *Australian Journal of Botany* 40(6): 887 - 896.
- Ikram N, Dawar S and Imtiaz F (2015). X rays exposure on leguminous seeds in combination with *Aerva javanica* parts powder for the promotion of growth and management of root rot fungal pathogens. *European Journal of Botany Plant Sciences and Pathology* 2: 1-10.
- Jeng TL, Shih YJ, Lai CC, Wu MT and Sung JM (2010). Ant-oxidative characterization of NaN_3 induced common bean mutants. *Food Chemistry* 119: 1006-1011.
- Jeng TL, Tseng TH, Wang CS, Chen CL and Sung JM (2003). Starch biosynthesizing enzymes in developing grains of rice cultivar Tainung 67 and its sodium azide-induced rice mutant. *Field Crops Research* 84: 261–269.
- Jeng TL, Tseng TH, Wang CS, Chen CL and Sung JM (2006). Yield and grain uniformity in contrasting rice genotypes suitable for different growth environments. *Field crop research* 99: 59-66.
- Jin-Hu Wu, Ross F, Brian, Murray, Yilin Jia, Paul, Datson and Jingli Z (2011). Induced polyploidy dramatically increases the size and alters the shape of fruit in *Actinidia chinensis*. *Annals of Botany* 109: 169-179.
- Khan A Irfan and Farook SA (1998). Mutation breeding in pulse crops. In: Nizam J, Khan IA, Farook SA (eds), *Genetics and Breeding Strategies for Improvement of Pulse Crops*. pp-104-125.
- Kharkwal MC (1983). Mutation breeding for chickpea improvement. *International Chickpea Newsletter* 9: 4-5.
- Kharkwal MC (2008). A brief history of plant mutagenesis. In: Shu QY, Forster BP, Nakagawa H, (eds). *Plant mutation Breeding and Biotechnology*. Wallingford: CABI; p. 21–30.
- Kiruki S, Onek LA and Limo M. (2006). Azide-based mutagenesis suppresses *Striga hermonthica* seed germination and parasitism on maize varieties. *African Journal of Biotechnology* 5: 866-870.

- Kumar G and Srivastava, P (2010). Comparative radiocytological effect of gamma rays and laser rays on safflower. *Romanian Journal of Biology - Plant Biology* 55(2): 105-111.
- Kumar P, Sharma V, Yadav P, Singh B (2017). Gamma ray irradiation for crop protection against salt stress. *Defence Life Science Journal* 2: 292-300.
- Kumar S (1988). Recessive monogenic mutation in grain pea (*Pisum sativum*) that causes pyridoxine requirement for growth and seed production. *Journal of Bioscience* 13: 415-418.
- Kurowska M, Pawlowska AL, Gnizda D, Maluszynski M and Szarejko I (2012) Molecular analysis of point mutations in a barley genome exposed to MNU and gamma rays. *Mutation Research* 738-739: 52-70.
- Leitao JM (2012). Chemical mutagenesis. In: Shu QY, Forster BP, Nakagawa H (eds). *Plant Mutation Breeding and Biotechnology* Wallingford: CABI. p. 135-158.
- Luan Y, Zhang J, Gao X and An L (2007). Mutation induced by ethylmethanesulphonate (EMS), in vitro screening for salt tolerance and plant regeneration of sweet potato (*Ipomoea batatas* L.). *Plant Cell Tissue and Organ Culture* 88:77-81.
- Lukanda TL, Kalonji-Mbuyi A, Nkongolo KKC and Kizungu RV (2013). Effect of gamma irradiation on morpho-agronomic characteristics of groundnut (*Arachis hypogaea* L.). *American Journal of Plant Sciences* 4: 2186-2192.
- Manawadu IP, Dahanayake N and Senanayake SGJN (2016). Colchicine Induced Tetraploids of Radish (*Raphanus sativus* L.). *Tropical Agricultural Research and Extension* 19: 173-183.
- Mba C, Afza R and Shu QY (2012). Mutagenic radiations: X-rays, ionizing particles and ultraviolet. In: Shu QY, Forster BP, Nakagawa H, (eds). *Plant Mutation Breeding and Biotechnology*. Wallingford: CABI; pp 83-90.
- Mba C, Afza R and Bado S (2010). Induced mutagenesis in plants using physical and chemical agents. In: Davey MR, Anthony P (eds). *Plant cell culture: essential methods*. Chichester: John Wiley & Sons, Ltd.; pp 111-130.
- Mba C (2013). Induced mutations unleash the potentials of plant genetic resources for food and agriculture. *Agronomy* 3(1): 200-231.
- Mensah JK and Obadoni B (2007). Effects of sodium azide on yield parameters of groundnut (*Arachis hypogaea* L.). *African Journal of Biotechnology* 6: 668-671.
- Molina-Cano JL, Simian JP, Sopena A, Perez-Vendrell AM, Dorsch S, Rubiales D, Swanston JS and Jahoor A (2003). Mildew-resistant mutants induced in North American two- and six-rowed malting barley cultivars. *Theoretical and Applied Genetics* 107: 1278-1287.
- Mondal S, Badigannavar AM, Kale DM and Murty GSS (2007). Induction of genetic variability in a disease resistant groundnut breeding line. *Newsletter, Founders day special issue* 285.
- Mosisa G, Muthuswamy M and Petros Y (2014). Effect of chemical mutagen through Hydroxylamine Hydrochloride on quantitative traits variation in *Phaseolus vulgaris*. *International Journal of Scientific and Technology Research* 3(2): 76-79
- Murali KM, Vanitha J, Jiang S and Ramachandran S (2013). *Molecular Plant Breeding* 4: 128-135.
- Nakata Y, Ueno M, Kihara J, Ichii M, Taketa S and Arase S (2008). Rice blast disease and susceptibility to pests in a silicon uptake-deficient mutant of rice. *Crop protection* 27: 865-868.
- Navabi Y, Norouzi M, Arab M and Daylami SD (2016). Mutagenesis via Exposure to Gamma-Rays in Tuberose (*Polianthes tuberosa*). *Electronic Journal of Biology* 12:168-172.
- Okubara PA, Anderson PA, Ochoa OE and Michelmore RW (1994). Mutant of downy mildew resistance in Lettuce (*Lactuca sativa*). *Genetics* 137: 867-874.
- Olsen O, Wang X and Von Wettstein D (1993). Sodium azide mutagenesis: preferential generation of A.T-->G.C transitions in the barley

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- ant18 gene. Proceedings of National Academics of Sciences 90: 8043-8047.
- Papadopoulou K, Melto RE, Leggett M, Daniels MJ and Osbourn AE (1996). Compromised disease resistance in saponin deficient plant. *Proceedings of the National Academy of Sciences USA* 96: 12923-12928.
- Prina AR and Favret EA (1983). Parabolic effect in sodium azide mutagenesis in barley. *Hereditas* 98: 89-94.
- Rafiq M, Kulmi M, Mogali SC, Patil KS and Leelavathi TM (2017). Isolation of High-Yielding Mutants through EMS-Induced Mutagenesis in Linseed (*Linum usitatissimum* L.). *International Journal of Current Microbiology and Applied Sciences* 6: 278-285.
- Rafiq M, Mali M, Naqvi SHA, Dahot MU, Faiza H and Khatari A (2012). *Pakistan Journal of Biotechnology* 9: 83-89.
- Rahman SM, Takagi Y, Kubota K, Miyamoto K and Kawakita T (1994). High Oleic Acid content in Soya bean. *Bioscience, Biotechnology and Biochemistry* 58: 1070-1072.
- Roy S and Biswas AK (2005). Isolation of a white flowered mutant through seed culture in *Spathoglottis plicata* Blume. *Cytologia* 70: 1-6.
- Samiullah K, Wani MR and Parveen K (2004). Induced genetic variability for quantitative traits in *Vigna radiate* (L) wilczek. *Pakistan Journal of Botany* 36: 845-850.
- Satoh H, Matsusaks H and Kumamaru T (1979). Use of N methyl N nitrosoarea treatment of fertilised egg cells for saturation mutagenesis of rice. *Breeding Science* 60: 475-485.
- Sinhamahapatra SP and Rakshit SC (1990). Response to selection for plant height in X-ray treated population of jute (*Corchorus capsularis* L.) cv. JRC 212. *Euphytica* 51: 95-99.
- Stadler L. 1930. The frequency of mutation of specific genes in maize. *Anatomical Record* 47: 381.
- Talebi AB and Shahrokhifar B (2012). Ethyl Methane Sulphonate induced mutagenesis in Malaysian Rice (cv. MR219) for lethal dose determination. *American journal of Plant Science* 3: 1661-1665.
- Venegas-Caleron M, Martinez-Force and Garces R (2008). Lipid characterization of a wrinkled sunflower mutant. *Phytochemistry* 69: 684-691.
- Wang L, Zhang B, Li J, Yang X and Ren Z. (2014). Ethyl Methanesulfonate (EMS)-Mediated Mutagenesis of Cucumber (*Cucumis sativus* L.). *Agricultural Sciences* 5: 716-721.
- Warghat AR, Rampure NH and Singh P (2011). Effect of sodium azide and gamma rays treatments on percentage germination, survival, morphological variation and chlorophyll mutation in musk okra (*Abelmoschus moschatus* L.). *International Journal of Pharmacy and Pharmaceutical Sciences* 3: 483-486.
- Weber S, Luhs W and Friedt W (2004). Plant Breeding Dept., IFZ - Research Centre for Biosystems, Land Resources & Nutrition, Justus-Liebig-University of Giessen, Heinrich-Buff-Ring 26-32, D-35392 Giessen, Germany.